

Synthesis of Photoactivatable Analogues of Lysophosphatidic Acid and Covalent Labeling of Plasma Proteins

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Lysophosphatidic acids bearing a benzophenone group in either the sn-1 or sn-2 chain of an oleoyl-type ester or oleyl-type ether chain and ³²P in the phosphate group were synthesized. The benzophenone moiety was introduced by selective hydroboration of the double bond of enyne 11 at low temperature, followed by a Suzuki reaction with 4-bromobenzophenone. The key intermediates for the preparation of esterlinked lysophosphatidic acid (LPA) 1 and 3 were obtained in one pot by a modified DIBAL-H reduction of orthoformate intermediate 22. These probes were shown to covalently modify a single protein target in rat plasma containing albumin and several protein targets in rat plasma containing a low level of albumin.

Introduction

Lysophosphatidic acid (LPA) has the simplest structure of natural phospholipids, yet it has attracted significant attention because of the diversity and importance of its biological effects.¹⁻⁵ LPA is not a single compound; rather, it comprises a family of related mediators that differ in the length and degree

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of unsaturation of a hydrocarbon chain linked to a glycerol backbone via ester,⁶ ether,⁷ or vinyl ether⁸ bonds at either the sn-1 or sn-2 position. Different LPA family members have distinct biological activities.9 LPA plays a key role in a myriad of physiological processes, including Ca2+ flux, vascular and neuronal function, cell growth/death, and cell migration.^{2,10} It is now appreciated that LPA has numerous intracellular, extracellular, and cell-surface targets. LPA exerts its activities mainly through several cell-surface G protein-coupled receptors (GPCR), including the widely reported LPA₁, LPA₂, and LPA₃

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SCHEME 1. Retrosynthetic Strategy

receptor subtypes. Recently, the LPA₄/GPR23/P2Y9 receptor¹¹ and the nuclear transcription factor PPAR γ^{12} were identified as LPA receptors. LPA also interacts with numerous binding proteins, including albumin¹³ and gelsolin.¹⁴ It has been speculated that interactions with binding proteins may modulate the biological activity of LPA.

Despite recent progress elucidating many of the biological properties of LPA, a detailed understanding of the key roles of LPA in many disease processes has not yet been achieved. To gain insights into the protein targets of LPA, we have prepared photoreactive analogues of LPA. Photoactivatable lipid analogues have been widely employed to study hydrophobic binding sites of lipid targets. 15 Recently, we reported the first synthesis and characterization of two photoactivatable analogues of sphingosine 1-phosphate (S1P).¹⁶ Likewise, photoactivatable analogues of bioactive glycerophospholipids¹⁷ and lysosphingophospholipids¹⁸ have been reported recently. One previous study reported the synthesis and characterization of a photoreactive LPA analogue containing a trifluoromethylphenyldiazirine group for labeling of fetal bovine serum and LPA responsive cells.¹⁹ In this paper, we report the syntheses of additional photoactivatable analogues of acyl- and O-alkyllinked LPAs in which the long chain resembles an oleoyl (ester) or oleyl (ether) chain, respectively. The rationale for this approach is that the oleoyl chain has been shown to represent the maximally effective fatty acyl chain of natural LPAs in previous structure-activity studies.²⁰ We have prepared both the 2-lyso (1 and 2) and 1-lyso (3 and 4) regioisomeric analogues of [32 P]-LPA, as well as the *O*-methoxymethyl (MOM)-protected photoreactive analogues of [32 P]-LPA (denoted as **20P** and **27P**). In the current analogues, benzophenone was employed as the photoprobe for labeling of proteins because it is readily activated with long-wavelength UV light, is stable in most of the chemical reactions used during the synthesis of the probe, and generally yields covalent products modified at a single site. In the four probes reported herein, the benzophenone moiety is linked to the end of a Δ^9 cis hydrocarbon chain. These analogues were characterized via the covalent modification of rat plasma proteins.

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Results and Discussion

Retrosynthetic Analysis. Our retrosynthetic strategy for the synthesis of 1 and 2 is outlined in Scheme 1. Analogues 1 and 2 were envisioned as being produced via enzymatic phosphorylation and deprotection of 2-MOM-protected 3-hydroxyglyceryl ester 5 (X = O) or ether ($X = H_2$), respectively. Compound 5 can be generated by the selective removal of the sn-3 protecting group (PG) from 6, whereas intermediate 6 is accessible from carboxylic acid 7 (X = O) or from the corresponding alcohol ($X = H_2$). The introduction of the benzophenone probe into 7 may be achieved by the regioselective addition of 9-BBN to the double bond in 8, provided that no reaction occurs at the internal triple bond, followed by Suzuki coupling with 4-bromobenzophenone. Compounds 3 and 4 can be prepared from the regioisomers of 5-8 using similar schemes.

Conversion of 2-Decyn-1-ol to Alcohol 14. Scheme 2 outlines the synthesis of Δ^9 alcohol 14. The acetylene zipper reaction of commercially available 2-decyn-1-ol²² gave terminal acetylide 9. After the hydroxy group was protected as a THP ether to give 10 in nearly quantitative yield, addition of allyl bromide to the lithium anion of 9 provided enyne 11 in 79% yield. The benzophenone moiety was introduced by selective hydroboration of the double bond of 11 at low temperature, without reaction of the organoborane with the triple bond, followed by the Suzuki reaction of the terminal boronate with 4-bromobenzophenone. After the THP group of 12 was re-

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SCHEME 2. Synthesis of Long-Chain Alcohol 14a

^a (a) LiNH(CH₂)₃NH₂, *t*-BuOK/H₂N(CH₂)₃NH₂; (b) DHP, PPTS, CH₂Cl₂; (c) (i) *n*-BuLi, THF, −78 °C to room temperature and (ii) allyl bromide, CuI, THF, −78 °C; (d) (i) 9-BBN, THF, −15 °C to room temperature and (ii) 4-bromobenzophenone, Pd(PPh₃)₄, K₃PO₄, THF/DMF, reflux, 2 h; (e) PPTS (cat.), MeOH; (f) H₂, Lindlar, MeOH.

SCHEME 3. Synthesis of Glyceride 20 via FMOC-Ester 18^a

^a (a) PDC/DMF, room temperature, 2 days; (b) (*R*)-(−)-2,2-dimethyl-1,3-dioxolane-4-methanol, DCC, DMAP, CH₂Cl₂, room temperature; (c) 0.4 N HCl/90% dioxane, room temperature; (d) FMOC−chloroformate, DMAP, CH₂Cl₂, −10 °C; (e) MOMCl, *i*-Pr₂NEt, CH₂Cl₂; (f) piperidine, CH₂Cl₂.

moved, Lindlar-catalyzed hydrogenation of the triple bond in 13 afforded *Z* alcohol 14 in very high yield.

Synthesis of Ester-Linked LPA Probes 1 and 3. Conversion of Alcohol 14 to Glyceride 20. Oxidation of alcohol 14 with PDC gave acid 15, which was converted to ester 16 by reaction with (*R*)-isopropylideneglycerol (Scheme 3). After removal of the acetonide protecting group in 16 afforded diol 17, monoacylation of the resultant primary hydroxy group was accomplished according to a reported procedure²³ with 1 equiv of FMOC—chloroformate, providing secondary alcohol 18 in 28% yield based on diol 17. This yield is comparable to that

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SCHEME 4. Synthesis of 1 and 3^a

 a (a) (i) HC(OMe)₃, CSA (cat.), CH₂Cl₂, room temperature and (ii) DIBAL-H, 0 °C; (b) **15**, PPh₃, DIAD, THF, 0 °C to room temperature; (c) CAN, CH₃CN/H₂O (6:1); (d) (i) DGK, [32 P]ATP, pH 6.6, 37 °C, 2 h and (ii) TMSBr, CH₂Cl₂, 0 °C.

reported with a similar diol.²³ Treatment of **18** with MOM–Cl provided glyceride **19**, and removal of the FMOC group (10% piperidine in CH₂Cl₂) afforded **20**. Although this multistep synthesis is straightforward, the overall yield of ester **20** is low and the method does not allow for the preparation of the regioisomer of **20**. To overcome these deficiencies, an alternative synthetic strategy was designed (see Scheme 4).

Syntheses of 1 and 3 via 3-O-PMP-sn-glycerol (21). Our revised synthetic scheme begins with the conversion of 3-O-PMP-sn-glycerol (21)²⁴ to glyceride 20 and its regioisomer 27 (Scheme 4). The MOM protecting group was introduced into diol 21 prior to ester formation by transesterification with trimethyl orthoformate using a modification of a previously reported procedure.²⁵ The key step in this scheme is the protection of one of the hydroxy groups of diol 21 as a MOM ether via orthoformate intermediate 22, followed by reduction with DIBAL-H in toluene. When the reduction was conducted at -78 °C, regioisomers 23 and 24 were obtained in a ratio of 23:1 and an overall yield of 96% after chromatography. Thus, this methodology is a highly efficient route to the precursor of 1-lyso-LPA product 3 but not to the 2-lyso-LPA product 1. We altered the reduction conditions to obtain an adequate amount of primary alcohol 24 for conversion to 1-acyl-LPA product 1, which is needed in the photolabeling experiments. When the DIBAL-H reduction was carried out at 0 °C, compound 24 was obtained in 14% yield, which sufficed for the preparation of 1. Esterification of 23 and 24 with acid 15 using Mitsunobu reaction conditions, followed by removal of the PMP group in 25 and 26 with CAN, provided 27 and 20. Compounds 1 and 3 were obtained by phosphorylation of 27 and 20 with

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Synthesis of Ether-Linked Probes 2 and 4^a

^a (a) (i) CH₃SO₂Cl, Et₃N, 0 °C to room temperature and (ii) LiBr, THF, reflux; (b) (i) (n-Bu)₂SnO, CHCl₃/MeOH (10:1), reflux and (ii) 28, CsF, 18-crown-6, DMF, room temperature, 1 day; (c) MOMCl, i-Pr₂NEt, CH₂Cl₂; (d) CAN, CH₃CN/H₂O (6:1); (e) (i) DGK, $[^{32}P]$ ATP and (ii) TMSBr, CH2Cl2, 0 °C.

diacylglycerol kinase (DGK) and [32P]ATP.26 The final step, deprotection of the MOM group of 20P and 27P, was accomplished in quantitative yield with trimethylsilyl bromide in dry CH₂Cl₂ at 0 °C.

Synthesis of Ether-Linked LPA Probes 2 and 4. Scheme 5 depicts the strategy we used to introduce an O-alkyl chain bearing a benzophenone probe into the sn-1 or sn-2 position of glycerol. Reaction of diol 21 with dibutyltin oxide afforded cyclic tin intermediate 29,27 which was used without purification. Ethers 30 and 31 were obtained in 80 and 10% yields, respectively, by nucleophilic substitution of bromide 28 with intermediate 29. After the hydroxy group in 30 or 31 was protected as a MOM ether, the PMP group was removed with CAN to give precursor 34 or 35 in high yield. Ether-linked probes 2 and 4 were obtained from 34 and 35 using the same phosphorylation and deprotection methodology as that described for the preparation of 1 and 3.

Plasma Protein Photolabeling. The goal of this research was to generate tools that can be used to distinguish targets of individual LPA family members, i.e., acyl vs O-alkyl and sn-1 vs sn-2 linked aliphatic benzophenones. To that end, we tested five of the synthesized target compounds and intermediates for the ability to photolabel plasma proteins. Probes 1 and 2 allow for a comparison of the sn-1 acyl vs O-alkyl analogues, whereas 2 and 4 allow for a comparison of the sn-1 vs sn-2 regioisomers. Comparisons between compounds 1 and 3 were not made because of facile acyl migration inherent in probe 3.28 However, the sn-3 phosphorylation products (20P and 27P) of synthetic intermediates 20 and 27 afford an additional comparison between the acyl regioisomers of LPA without the risk of acyl migration. Plasma from the Sprague-Dawley rat ("normal rat plasma" (NRP)) contains 19.4 mg of albumin per milliliter²⁹ and was used as a surrogate for normal human plasma. Conversely, plasma from the Nagase analbuminemic rat ("analbuminemic rat plasma" (ARP)) contains only 0.4 mg of albumin per milliliter.²⁹ This material was used as a tool to examine binding to plasma proteins other than albumin.

Benzophenone-containing LPA probes were characterized by UV-dependent labeling of plasma proteins (Figure 1). Probes 1, 2, 4, 20P, and 27P all showed time- (Figure 1A) and concentration-dependent (data not shown) labeling of predominantly one protein band in NRP. On the basis of the fact that LPA was initially purified from serum as biological activity associated with albumin¹³ and the size of the labeled band in this study (\sim 70 kDa), we conclude that this protein is likely albumin. In contrast, four or five targets were identified on photolabeling of plasma with a low content of albumin (ARP) with the same probes (Figure 1B). Once again, labeling for all probes was time (Figure 1B) and dose (data not shown) dependent. The intensity of the individual labeled bands varied among the probes, but all of the probes modify similar targets in both NRP and ARP. This was expected because these targets are presumed to be transport proteins in which the interactions are based more on hydrophobicity than on stereochemistry or regiochemistry. Further identification and characterization of plasma protein targets of LPA-containing benzophenones, as well as examination of the pharmacology of these reagents at individual LPA GPCR and cellular targets, is currently underway in our laboratories.

Experimental Section

Tetrahvdro-2-(tridec-12-en-9-vnvloxv)-2H-pvran (11). To a solution of alkyne 10 (2.00 g, 8.40 mmol) in 20 mL of dry THF was added n-BuLi (3.5 mL, 10.1 mmol, a 2.89 M solution in hexane) at -78 °C under N_2 . After being stirred for 1 h at -78°C, the reaction mixture was warmed to room temperature and stirred for an additional 1.5 h. The mixture was then slowly added to a solution of allyl bromide (1.22 g, 10.1 mmol) and CuI (100 mg, 0.50 mmol) in 20 mL of THF at −78 °C. After the reaction mixture was stirred overnight at room temperature, the mixture was filtered through a short silica gel column. The solvent was removed, and the residue was purified by chromatography (hexane/EtOAc 20:1) to afford 11 (1.85 g, 79%) as a colorless oil: ¹H NMR δ 1.24-1.90 (m, 18H), 2.14-2.22 (m, 2H), 2.90-2.97 (m, 2H), 3.34-3.41 (m, 1H), 3.48-3.52 (m, 1H), 3.69-3.76 (m, 1H), 3.84-3.89 (m, 1H), 4.55-4.59 (m, 1H), 5.06-5.11 (m, 1H), 5.27-5.36 (m, 1H), 5.77–5.84 (m, 1H); 13 C NMR δ 18.7, 19.6, 23.1, 25.4, 26.1, 28.8, 29.0, 29.0, 29.3, 29.7, 30.7, 62.2, 67.6, 76.4, 82.8, 98.8, 115.5, 133.3; HR-MS [MNa⁺] m/z calcd for $C_{18}H_{30}O_2Na$ 301.2138, found 301.2107.

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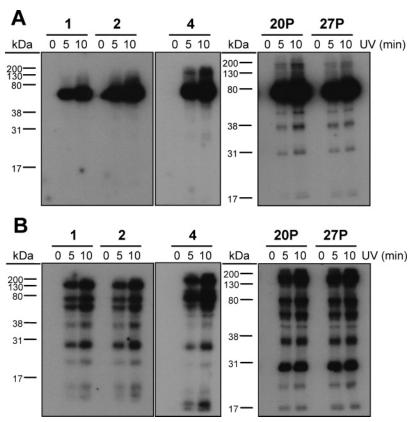


FIGURE 1. Covalent modification of plasma proteins by benzophenone-containing LPA analogues. Plasma (5% in PBS) from Sprague—Dawley (19.4 mg albumin/mL) (**A**) or Nagase rats (0.4 mg albumin/mL) (**B**) was incubated with compounds **1**, **2**, **4**, **20P**, and **27P** (50 nM, DMSO 10% final concentration) in the dark for 30 min prior to UV irradiation for up to 10 min on ice. Samples were diluted with 2 × sample loading buffer and separated by SDS—PAGE. Gels were subsequently dried, and radioactive bands were visualized by autoradiography.

4-(13-(Tetrahydro-2H-pyran-2-yloxy)tridec-4-ynyl)benzophenone (12). A flask charged with 11 (1.11 g, 4.00 mmol) and 15 mL of THF was cooled to -15 °C in a salt-ice bath, and 9-BBN (12 mL, 6.0 mmol, a 0.5 M solution in THF) was added dropwise under N₂. After the addition, the reaction mixture was warmed to room temperature and stirred for 5 h. 4-Bromobenzophenone (1.04 g, 4.00 mmol), Pd(PPh₃)₄ (230 mg, 0.20 mmol), DMF (15 mL), and anhydrous K₃PO₄ (1.57 g, 8.00 mmol) were added, and the resulting mixture was heated at reflux for 2 h. The reaction mixture was poured into 100 mL of water and extracted with CH₂Cl₂ (4 × 40 mL). The combined organic layers were washed with 100 mL of water and dried (Na₂SO₄). The solvent was removed under vacuum, and the residue was purified by chromatography (hexane/EtOAc 8:1) to provide 12 (1.21 g, 66%) as a colorless oil: ¹H NMR δ 1.28-1.90 (m, 20H), 2.12-2.22 (m, 4H), 2.80 (t, 2H, J = 7.6 Hz), 3.32-3.40 (m, 1H), 3.43-3.50 (m, 1H), 3.68-3.74 (m, 1H), 3.82-3.87 (m, 1H), 4.53-4.58 (m, 1H), 7.28-7.80 (m, 9H). ¹³C NMR δ 18.0, 18.5, 19.5, 25.3, 26.0, 28.6, 28.9, 29.2, 29.5, 30.2, 30.6, 34.6, 62.1, 67.4, 77.2, 79.1, 80.9, 98.6, 128.0, 128.3, 129.7, 130.1, 132.0, 135.1, 137.7, 146.9, 196.1; HR-MS [MNa⁺] m/z calcd for C₃₁H₄₀O₃Na 483.2870, found 483.2880.

4-(13-Hydroxytridec-4-ynyl)benzophenone (13). A solution of **12** (370 mg, 0.80 mmol) and a catalytic amount of PPTS in MeOH was heated at reflux for 3 h. Concentration, followed by purification of the residue by chromatography (hexane/EtOAc 4:1), provided **13** (282 mg, 93%): 1 H NMR δ 1.25–1.60 (m, 12H), 1.80–1.85 (m, 2H), 2.14–2.21 (m, 4H), 2.75 (s, 1H), 2.80 (t, 2H, J = 7.6 Hz), 3.58 (t, 2H, J = 6.8 Hz), 7.27–7.79 (m, 9H); 13 C NMR δ 17.9, 18.5, 25.5, 28.5, 28.8, 28.9, 29.1, 30.0, 32.4, 34.5, 62.3, 79.1, 80.9, 127.9, 128.2, 129.6, 130.1, 132.0, 134.9, 137.5, 146.9, 196.2; HR-MS [MNa⁺] m/z calcd for C₂₆H₃₂O₂Na 399.2294, found 399.2285.

4-((*Z*)-**13-Hydroxytridec-4-enyl)benzophenone** (**14**). Lindlar catalyst (15 mg, 5.5 wt %) was added to a solution of **13** (270 mg,

0.72 mmol) in 10 mL of MeOH. The mixture was stirred under a hydrogen balloon until the starting alkyne was consumed (monitored by thin-layer chromatography (TLC), hexane/EtOAc 2:1). The solvent was removed under vacuum, and the residue was purified by chromatography (hexane/EtOAc 4:1) to give **14** (262 mg, 97%) as an oil: $^1\mathrm{H}$ NMR δ 1.25–1.48 (m, 10H), 1.50–1.56 (m, 2H), 1.68–1.74 (m, 2H), 1.96–2.13 (m, 4H), 2.45 (s, 1H), 2.69 (t, 2H, J=7.6 Hz), 3.59 (t, 2H, J=6.8 Hz), 5.33–5.40 (m, 2H), 7.25–7.77 (m, 9H); $^{13}\mathrm{C}$ NMR δ 25.6, 26.5, 27.1, 29.0, 29.2, 29.3, 29.5, 30.9, 32.5, 35.2, 62.5, 128.0, 128.2, 128.7, 129.7, 130.1, 130.5, 132.0, 134.9, 137.6, 147.7, 196.4; HR-MS [MNa⁺] m/z calcd for $\mathrm{C}_{26}\mathrm{H}_{34}\mathrm{O}_{2}\mathrm{Na}$ 401.2451, found 401.2454.

(*Z*)-13-(4-Benzoylphenyl)-9-tridecenoic Acid (15). A solution of 14 (730 mg, 1.93 mmol) and PDC (7.3 g, 19.3 mmol) in DMF (20 mL) was stirred at room temperature for 2 days. Water (100 mL) was added, and the mixture was extracted with CH₂Cl₂ (3 × 50 mL). The combined organic layers were washed with water (100 mL) and dried (Na₂SO₄). The solvent was removed under vacuum, and the residue was purified by chromatography (hexane/EtOAc 2:1) to give 15 (657 mg, 87%) as a viscous oil: ¹H NMR δ 1.24–1.40 (m, 8H), 1.58–1.75 (m, 4H), 1.96–2.12 (m, 4H), 2.33 (t, 2H, *J* = 7.2 Hz), 2.70 (t, 2H, *J* = 7.6 Hz), 5.33–5.44 (m, 2H), 7.26–7.79 (m, 9H); ¹³C NMR δ 24.6, 26.7, 27.2, 28.9, 29.0, 29.1, 29.5, 31.0, 34.0, 35.4, 128.1, 128.3, 128.9, 129.9, 130.3, 130.6, 132.1, 135.0, 137.8, 147.8, 179.9, 196.5; HR-MS [MNa⁺] m/z calcd for C₂₆H₃₂O₃Na 415.2244, found 415.2255.

1-*O*-(**13**-(**4**-Benzoylphenyl)-**9**-(**Z**)-tridecenoyl)-**3**-*O*-FMOC-snglycerol (**18**). DCC (6.3 mg, 0.031 mmol) and DMAP (1.0 mg, 8.1 μ mol) were added to a solution of **15** (10.0 mg, 0.025 mmol) and (*R*)-(-)-2,2-dimethyl-1,3-dioxolane-4-methanol (4.0 mg, 0.031 mmol) in 1 mL of CH₂Cl₂. After the mixture was stirred at room temperature for 40 h, the solvent was removed under vacuum, and the residue was purified by chromatography (hexane/EtOAc 4:1) to provide (*Z*)-13-benzoylphenyl-9-tridecenoic acid (*S*)-(2,2-di-

methyl-1,3-dioxolan-4-yl)methyl ester (**16**) (11.9 mg, 92%): $^{1}\mathrm{H}$ NMR δ 1.23–1.37 (m, 8H), 1.37 (s, 3H), 1.43 (s, 3H), 1.60–1.75 (m, 4H), 1.96–2.12 (m, 4H), 2.34 (t, 2H, J=7.6 Hz), 2.70 (t, 2H, J=7.6 Hz), 3.71–3.75 (m, 1H), 4.05–4.18 (m, 3H), 4.28–4.33 (m, 1H), 5.33–5.42 (m, 2H), 7.26–7.80 (m, 9H); $^{13}\mathrm{C}$ NMR δ 24.8, 25.3, 26.6, 26.7, 27.1, 29.0, 19.0, 29.0, 29.5, 31.0, 34.0, 35.4, 64.4, 66.2, 73.5, 109.7, 128.1, 128.4, 128.9, 129.8, 130.2, 130.5, 132.1, 135.0, 137.8, 147.7, 173.5, 196.3.

One drop of concentrated HCl was added to a solution of **16** (90 mg, 0.18 mmol) in 90% dioxane (3 mL). After the mixture was stirred at room temperature for 4 h, the solvent was removed under vacuum and the residue was purified by chromatography (hexane/EtOAc 1:1) to give 1-((*Z*)-13-(4-benzoylphenyl)-9-tridecenoyl))-*sn*-glycerol (**17**) (60 mg, 73%): ¹H NMR δ 1.23–1.38 (m, 8H), 1.55–1.68 (m, 4H), 1.95–2.12 (m, 4H), 2.33 (t, 2H, J = 7.6 Hz), 2.70 (t, 2H, J = 7.6 Hz), 2.75 (br s, 2H), 3.55–3.68 (m, 2H), 3.88–3.94 (m, 1H), 4.15 (t, 2H, J = 4.8 Hz), 5.34–5.43 (m, 2H), 7.25–7.79 (m, 9H); ¹³C NMR δ 24.8, 26.6, 27.1, 29.0, 29.0, 29.0, 29.5, 31.0, 34.0, 35.4, 63.3, 65.0, 70.1, 128.2, 128.3, 128.9, 129.9, 130.3, 130.6, 132.2, 135.0, 137.8, 147.9, 174.2, 196.7.

A solution of **17** (58 mg, 0.12 mmol) and DMAP (15 mg, 0.12 mmol) in 1 mL of dry CH_2Cl_2 was cooled to -10 °C in a salt-ice bath. A solution of FMOC-chloroformate (32 mg, 0.12 mmol) in 2 mL of dry CH₂Cl₂ was added dropwise. After the reaction mixture was stirred for 30 min, the solution was washed with water (5 mL) and dried (Na₂SO₄). A small amount (12 mg) of starting material 17 was recovered after chromatography, and 18 (19 mg, 28%) was also obtained after chromatography (hexane/EtOAc 1:3): ¹H NMR δ 1.22–1.76 (m, 12H), 1.95–2.21 (m, 5H), 2.35 (t, 2H, J = 7.6Hz), 2.70 (t, 2H, J = 7.6 Hz), 4.10–4.28 (m, 5H), 4.41 (d, 2H, J= 7.2 Hz), 5.33-5.42 (m, 2H), 7.24-7.80 (m, 17H); 13 C NMR δ 24.8, 26.7, 27.2, 29.1, 29.1, 29.6, 30.9, 31.1, 34.1, 35.5, 46.7, 64.8, 68.1, 68.5, 70.1, 120.1, 125.1, 127.2, 127.9, 128.2, 129.0, 129.9, 130.3, 130.6, 132.2, 135.1, 137.9, 141.3, 143.2, 147.8, 155.2, 173.8, 196.6, 207.0; HR-MS [MNa⁺] m/z calcd for C₄₄H₄₈O₇Na 711.3292, found 711.3302.

1-O-(13-(4-Benzoylphenyl)-9-(Z)-tridecenoyl)-2-O-methoxymethyl-3-O-FMOC-sn-glycerol (19). Chloromethyl methyl ether $(7 \text{ mg}, 87 \,\mu\text{mol})$ was added to a stirred solution of **18** (6 mg, 8.7) μmol) and i-Pr₂NEt (11 mg, 87 μmol) in dry CH₂Cl₂ (1 mL) under N₂ at 0 °C. After the mixture was stirred at 0 °C for 2 h and at room temperature for 2 days, water (5 mL) and CH₂Cl₂ (5 mL) were added. The organic layer was washed with brine (5 mL) and water (5 mL) and then dried (Na₂SO₄). Product 19 (4.2 mg, 66% yield) was obtained after removal of the solvent and purification by chromatography (hexane/EtOAc 4:1): ¹H NMR δ 1.21–1.39 (m, 12H), 1.57–1.75 (m, 4H), 1.95–2.12 (m, 4H), 2.34 (t, 2H, J = 7.6 Hz), 2.70 (t, 2H, J = 7.6 Hz), 3.40 (s, 3H), 4.03–4.33 (m, 5H), 4.41 (d, 2H, J = 7.1 Hz), 4.73 (s, 2H), 5.33–5.44 (m, 2H), 7.24–7.83 (m, 17H); 13 C NMR δ 24.9, 26.8, 27.3, 29.1, 29.6, 31.1, 34.1, 35.5, 46.7, 55.7, 63.0, 67.0, 70.0, 72.4, 96.0, 100.0, 120.1, 125.1, 127.2, 127.9, 128.2, 128.4, 129.0, 130.0, 130.3, 130.7, 132.2, 135.1, 137.9, 141.3, 143.3, 147.8, 155.0, 173.4, 196.5; HR-MS [MNa⁺] m/z calcd for C₄₆H₅₂O₈Na 755.3560, found 755.3598.

1-O-(13-(4-Benzoylphenyl)-9-(Z)-tridecenoyl)-2-O-methoxymethyl-sn-glycerol (20) from 19. Piperidine (0.2 mL) was added to a solution of 19 (1.3 mg, 1.7 μ mol) in dry CH₂Cl₂ (2 mL) under nitrogen. The mixture was stirred at room temperature until the starting material was consumed (TLC, hexane/EtOAc 4:1). Concentration and purification by chromatography (hexane/EtOAc 2:1) afforded 20 (0.6 mg, 66%). The R_f value and the 1 H NMR spectrum were identical with data obtained for the same compound prepared from 26 (see below).

1-(4-Methoxyphenoxy)-3-(methoxymethoxy)propan-(2R)-2-ol (23) and 3-(4-Methoxyphenoxy)-(2R)-2-(methoxymethoxy)propan-1-ol (24). Trimethyl orthoformate (0.66 mL, 6.0 mmol) was added to a stirred suspension of diol 21 (594 mg, 3.0 mmol) and D-10-camphorsulfonic acid (12 mg, 50 μ mol) in 36 mL of dry CH₂Cl₂ at room temperature under nitrogen. The reaction mixture

was stirred at room temperature until diol 21 was fully consumed (TLC, hexane/EtOAc 1:1). The mixture was cooled to 0 °C, and 12 mL of DIBAL-H (18 mmol, a 1.5 M solution in toluene) was added slowly. Stirring was continued at 0 °C until the ortho ester intermediate disappeared (TLC, hexane/EtOAc 4:1). A solution of potassium sodium tartrate (10.2 g, 36.0 mmol) in water (40 mL) was added cautiously to quench the reaction, and the resulting mixture was stirred at room temperature overnight. The organic layer was separated, and the aqueous layer was extracted with CH2- Cl_2 (3 × 30 mL). The combined organic extracts were washed with water (50 mL) and dried (Na₂SO₄ and a small amount of K₂CO₃). The solvent was removed under vacuum, and the residue was purified by chromatography (a gradient of hexane/EtOAc from 10:1 to 1:1) to provide 603 mg (83%) of 23 and 99 mg (14%) of 24. Data for compound 23: ${}^{1}H$ NMR δ 3.31 (s, 1H), 3.35 (s, 3H), 3.65-3.77 (m, 5H), 3.96 (d, 2H, J = 5.8 Hz), 4.08-4.15 (m, 1H), 4.65 (s, 2H), 6.77-6.86 (m, 4H); 13 C NMR δ 55.0, 55.3, 68.9, 69.2, 69.4, 96.6, 114.4, 115.3, 152.5, 153.8; HR-MS [MNa⁺] m/z calcd for C₁₂H₁₈O₅Na 265.1046, found 265.1049. Data for compound **24**: ¹H NMR δ 2.69 (s, 1H), 3.43 (s, 3H), 3.71–3.86 (m, 5H), 3.93-4.04 (m, 3H), 4.79 (s, 2H), 6.78-6.87 (m, 4H); ¹³C NMR δ 55.6, 62.9, 68.3, 78.0, 96.7, 114.6, 115.4, 152.6, 154.0; HR-MS [MNa⁺] m/z calcd for $C_{26}H_{34}O_2Na$ 265.1046, found 265, 1037.

1-O-Methoxymethyl-2-O-(13-(4-benzoylphenyl)-9-(Z)-tridecenoyl)-3-*O*-(4-methoxyphenyl)-s*n*-glycerol (25). DIAD (10 mg, 49.5 μ mol) was added to a solution of acid 15 (16 mg, 41.3 μ mol), alcohol 23 (10 mg, 41.3 μ mol), and PPh₃ (13 mg, 49.5 μ mol) in dry THF (2 mL) at 0 °C. After the mixture was warmed to room temperature and stirred for 24 h, water (5 mL) and CH₂Cl₂ (5 mL) were added. The aqueous layer was extracted with CH_2Cl_2 (3 × 5 mL), and the combined organic layers were washed twice with water (5 mL) and dried (Na₂SO₄). After the solvents were removed under vacuum, the residue was purified by chromatography (hexane/ EtOAc 4:1) to give **25** (19 mg, 76%): ¹H NMR δ 1.24–1.36 (m, 8H), 1.58-1.76 (m, 4H), 1.95-2.12 (m, 4H), 2.35 (t, 2H, J=7.6Hz), 2.70 (t, 2H, J = 7.6 Hz), 3.34 (s, 3H), 3.76 (s, 3H), 3.79 (d, 2H), 4.06-4.13 (m, 2H), 4.64 (s, 2H), 5.29-5.35 (m, 1H), 5.36-5.44 (m, 2H), 6.80–6.87 (m, 4H), 7.27–7.82 (m, 9H); ¹³C NMR δ 24.9, 26.8, 27.3, 29.0, 29.1, 29.2, 29.6, 31.1, 34.3, 35.5, 55.3, 55.7, 65.8, 67.0, 70.8, 96.5, 114.6, 115.6, 128.2, 128.4, 129.0, 130.0,130.4, 130.7, 132.2, 135.1, 137.9, 147.8, 152.7, 154.1, 173.3, 196.5; HR-MS [MNa⁺] m/z calcd for $C_{38}H_{48}O_7Na$ 639.3292, found 639.3298.

1-*O*-(13-(4-Benzoylphenyl)-9-(*Z*)-tridecenoyl)-2-*O*-methoxymethyl-3-*O*-(4-methoxyphenyl)-sn-glycerol (26). Compound 26 was prepared in 78% yield according to the procedure used to prepare compound 25: 1 H NMR δ 1.24–1.35 (m, 8H), 1.58–1.78 (m, 4H), 1.95–2.13 (m, 4H), 2.30–2.35 (m, 2H), 2.68–2.73 (m, 2H), 3.41 (s, 3H), 3.76 (s, 3H), 4.02 (d, 2H, J = 5.3 Hz), 4.11–4.17 (m, 1H), 4.22–4.28 (m, 1H), 4.33–4.38 (m, 1H), 4.77 (s, 2H), 5.36–5.42 (m, 2H), 6.80–6.86 (m, 4H), 7.26–7.82 (m, 9H); 13 C NMR δ 24.9, 26.8, 27.2, 29.1, 29.1, 29.2, 29.6, 31.1, 34.1, 35.5, 55.6, 55.7, 63.6, 68.2, 73.3, 96.1, 114.6, 115.5, 128.2, 128.3, 129.0, 129.9, 130.3, 130.6, 132.1, 135.1, 137.9, 147.8, 152.7, 154.1, 173.5, 196.5; HR-MS [MNa⁺] m/z calcd for $C_{38}H_{48}O_7Na$ 639.3292, found 639.3297.

1-O-Methoxymethyl-2-O-(13-(4-benzoylphenyl)-9-(Z)-tridecenoyl)-sn-glycerol (27). CAN (181 mg, 0.33 mmol) was added to a solution of **25** (85 mg, 0.14 mmol) in 7 mL of CH₃CN/water (6:1) at room temperature. The mixture was stirred until compound **25** was consumed (\sim 1 h, TLC, hexane/EtOAc 1:1) and was then diluted with water (10 mL) and CH₂Cl₂ (10 mL). The aqueous layer was extracted with CH₂Cl₂ (3 × 6 mL), and the combined organic extracts were washed with brine (10 mL), dried (Na₂SO₄), filtered, and concentrated. The residue was purified by chromatography (hexane/EtOAc 2:1) to furnish **27** (49 mg, 69%): ¹H NMR δ 1.23–1.38 (m, 8H), 1.57–1.77 (m, 4H), 1.96–2.13 (m, 4H), 2.35 (t, 2H, J = 7.6 Hz), 2.70 (t, 2H, J = 7.6 Hz), 3.36 (s, 3H), 3.70–3.83 (m,

4H), 4.63 (s, 2H), 5.01–5.07 (m, 1H), 5.34–5.45 (m, 2H), 7.27–7.81 (m, 9H); $^{13}\mathrm{C}$ NMR δ 24.9, 26.7, 27.2, 29.1, 29.1, 29.2, 29.6, 31.1, 34.3, 35.5, 55.4, 62.3, 66.3, 73.1, 96.6, 128.2, 128.4, 129.0, 130.0, 130.4, 130.6, 132.2, 135.1, 137.9, 147.9, 173.6, 196.6; HR-MS [MNa⁺] m/z calcd for $\mathrm{C_{31}H_{42}O_6Na}$ 533.2874, found 533.2865.

1-*O***-(13-(4-Benzoylphenyl)-9-(***Z***)-tridecenoyl)-2-***O***-methoxymethyl-***sn***-glycerol (20) from Compound 26.** Compound **20** was prepared in 73% yield by the same procedure used to prepare compound **27**: 1 H NMR δ 1.24–1.38 (m, 8H), 1.56–1.76 (m, 4H), 1.95–2.17 (m, 4H), 2.33 (t, 2H, J = 7.6 Hz), 2.71 (t, 2H, J = 7.6 Hz), 3.42 (s, 3H), 3.58–3.71 (m, 2H), 3.80–3.85 (m, 1H), 4.14–4.24 (m, 2H), 4.74 (s, 2H), 5.35–5.44 (m, 2H), 7.24–7.82 (m, 9H); 13 C NMR δ 24.9, 26.7, 27.2, 29.1, 29.1, 29.6, 31.1, 34.1, 35.5, 55.7, 62.6, 63.2, 77.7, 96.6, 128.2, 128.3, 129.0, 129.9, 130.3, 130.6, 132.2, 135.1, 137.9, 147.8, 173.7, 196.5; HR-MS [MNa⁺] m/z calcd for C₃₁H₄₂O₆Na 533.2874, found 533.2863.

4-((Z)-Bromotridec-4-enyl)benzophenone (28). To 3 mL of CH₂Cl₂ under N₂ were added alcohol **14** (108 mg, 0.29 mmol) and Et₃N (43 mg, 0.43 mmol) at 0 °C. After the mixture was stirred for 5 min, methanesulfonyl chloride (49 mg, 0.43 mmol) was added, and the resulting reaction mixture was stirred at room temperature overnight, quenched with 5 mL of water, and extracted with CH₂- Cl_2 (3 × 5 mL). The combined organic phases were washed with brine, dried (Na₂SO₄), filtered, and concentrated. The crude mesylate was dissolved in dry THF (5 mL), and lithium bromide (50 mg, 0.57 mmol) was added. The reaction mixture was heated at reflux overnight and then cooled to room temperature and poured into water (5 mL) and CH₂Cl₂ (5 mL). The aqueous phase was extracted with CH₂Cl₂ (3 × 5 mL). The combined organic phases were washed with water (10 mL), dried (Na₂SO₄), and concentrated. The resulting yellow oil was purified by chromatography (hexane/EtOAc 10:1) to provide **28** (113 mg, 90%) as a colorless oil: ¹H NMR δ 1.22-1.45 (m, 10H), 1.68-1.86 (m, 4H), 1.96-2.12 (m, 4H), 2.70 (t, 2H, J = 7.8 Hz), 3.37 (t, 2H, J = 6.8 Hz), 5.35 - 5.44 (m, 2H),7.25–7.82 (m, 9H); 13 C NMR δ 26.6, 27.1, 28.0, 28.6, 29.1, 29.2, 29.5, 31.0, 32.7, 33.9, 35.4, 128.1, 128.2, 128.9, 129.8, 130.2, 130.5, 132.0, 135.0, 137.8, 147.7, 196.2; HR-MS [MNa⁺] m/z calcd for C₂₆H₃₃BrONa 463.1607, found 463.1600.

1-*O*-(13-(4-Benzovlphenyl)-9-(*Z*)-tridecenyl)-3-*O*-(4-methoxyphenyl)-sn-glycerol (30) and 2-O-(13-(4-Benzoylphenyl)-9-(Z)-tridecenyl)-3-O-(4-methoxyphenyl)-sn-glycerol (31). Di-nbutyltin oxide (64 mg, 0.26 mmol) was added to a solution of diol 21 (51 mg, 0.26 mmol) in 3 mL of CHCl₃/MeOH (10:1). After the resulting suspension was heated at reflux for 3 h to give a clear solution, the solvents were evaporated to give cyclic stannoxane intermediate 29 as a white solid. Cesium fluoride (78 mg, 0.51 mmol) and 18-crown-6 (8 mg, 26 μ mol) were added, and the solid mixture was dried overnight under high vacuum. After DMF (3 mL) and bromide 28 (71 mg, 0.16 mmol) were added, the reaction mixture was stirred at room temperature for 1 day. Water (0.5 mL) and EtOAc (10 mL) were added, and the mixture was stirred for an additional 1 h and was subsequently filtered through a pad of silica gel to remove an insoluble byproduct. The filtrate was washed with water (5 mL), dried (Na₂SO₄), and concentrated. The residue was purified by chromatography (hexane/EtOAc 2:1) to provide **30** (72 mg, 80%) and **31** (8.7 mg, 10%). Data for compound **30**: 1 H NMR δ 1.21–1.38 (m, 8H), 1.52–1.76 (m, 4H), 1.94–2.12 (m, 4H), 2.54 (br s, 1H), 2.70 (t, 2H, J = 7.6 Hz), 3.43-3.64 (m, 4H), 3.75 (s, 3H), 3.93-4.01 (m, 2H), 4.09-4.16 (m, 1H), 5.33-5.45 (m, 2H), 6.79–6.88 (m, 4H), 7.25–7.82 (m, 9H); ¹³C NMR δ 26.0, 26.7, 27.2, 29.2, 29.4, 29.4, 29.5, 29.6, 31.0, 35.4, 55.6, 69.1, 69.7, 71.5, 71.7, 114.6, 115.5, 128.1, 128.3, 128.9, 129.9, 130.3, 130.7, 132.1, 135.1, 137.9, 147.8, 152.7, 154.0, 196.4; HR-MS [MNa⁺] m/z calcd for C₃₆H₄₆O₅Na 581.3237, found 581.3249. Data for compound **31**: ${}^{1}H$ NMR δ 1.22–1.38 (m, 8H), 1.52– 1.77 (m, 4H), 1.95 - 2.14 (m, 4H), 2.68 (t, 2H, J = 7.8 Hz), 3.52 -3.86 (m, 5H), 3.76 (s, 3H), 3.94-4.04 (m, 2H), 5.34-5.45 (m, 2H), 6.79–6.91 (m, 4H), 7.25–7.81 (m, 9H); 13 C NMR δ 26.1, 26.8, 27.3, 29.3, 29.5, 29.5, 29.7, 30.1, 31.1, 35.5, 55.7, 62.5, 68.1, 70.7, 78.2, 114.7, 115.5, 128.2, 128.4, 129.0, 130.0, 130.4, 130.8, 132.2, 135.1, 138.0, 147.9, 152.8, 154.0, 196.6; HR-MS [MNa⁺] m/z calcd for $\rm C_{36}H_{46}O_5Na$ 581.3237, found 581.3252.

1-O-(13-(4-Benzoylphenyl)-9-(Z)-tridecenyl)-2-O-methoxymethyl-3-*O*-(4-methoxyphenyl)-s*n*-glycerol (32). Chloromethyl methyl ether (40 mg, 497 μ mol) was added under N₂ to a stirred solution of 30 (56 mg, 99 μ mol) and *i*-Pr₂NEt (64 mg, 497 μ mol) in dry CH₂Cl₂ (3 mL) at 0 °C. The reaction mixture was stirred at room temperature for 2 days. The solvent was removed, and the residue was purified by chromatography (hexane/EtOAc 4:1) to afford unreacted 30 (10 mg) and 32 (45 mg, 91%): ^{1}H NMR δ 1.22-1.38 (m, 8H), 1.52-1.76 (m, 4H), 1.96-2.15 (m, 4H), 2.70 (t, 2H, J = 7.8 Hz), 3.41 (s, 3H), 3.41–3.50 (m, 2H), 3.56–3.65 (m, 2H), 3.75 (s, 3H), 3.99-4.11 (m, 3H), 4.79 (s, 3H), 5.34-5.45 (m, 2H), 6.79-6.87 (m, 4H), 7.28 (d, 2H, J = 8.3 Hz), 7.47(t, 2H, J = 7.8 Hz), 7.57 (t, 1H, J = 7.3 Hz), 7.72 - 7.81 (m, 4H); 13 C NMR δ 26.0, 26.7, 27.2, 29.2, 29.4, 29.4, 29.6, 29.6, 31.1, 35.4, 55.4, 55.6, 68.7, 70.4, 71.7, 84.5, 96.2, 114.5, 115.4, 128.1, 128.3, 128.9, 129.9, 130.3, 130.7, 132.1, 135.1, 137.9, 147.8, 152.9, 153.8, 196.4; HR-MS [MNa⁺] m/z calcd for $C_{38}H_{50}O_6Na$ 625.3500, found 625.3519.

1-*O*-Methoxymethyl-2-*O*-(13-(4-benzoylphenyl)-9-(*Z*)-tridecenyl)-3-*O*-(4-methoxyphenyl)-sn-glycerol (33). Compound 33 was prepared in 89% yield from 31 by the same procedure used to prepare 32: 1 H NMR δ 1.22 $^{-}$ 1.39 (m, 8H), 1.55 $^{-}$ 1.62 (m, 2H), 1.68 $^{-}$ 1.76 (m, 2H), 1.97 $^{-}$ 2.04 (m, 2H), 2.07 $^{-}$ 2.13 (m, 2H), 2.70 (t, 2H, J = 7.8 Hz), 3.34 (s, 3H), 3.58 $^{-}$ 3.80 (m, 5H), 3.76 (s, 3H), 3.97 $^{-}$ 4.08 (m, 2H), 4.65 (s, 2H), 5.34 $^{-}$ 5.44 (m, 2H), 6.80 $^{-}$ 6.88 (m, 2H), 7.26 $^{-}$ 7.81 (m, 9H); 13 C NMR δ 26.0, 26.7, 27.3, 29.3, 29.4, 29.5, 29.7, 30.0, 31.1, 35.5, 55.2, 55.7, 66.9, 68.2, 70.7, 77.1, 96.7, 114.6, 115.5, 128.2, 128.3, 128.9, 129.9, 130.3, 130.8, 132.1, 135.1, 137.9, 147.8, 153.0, 153.9, 196.5; HR-MS [MNa⁺] m/z calcd for $C_{38}H_{50}O_6$ Na 625.3500, found 625.3497.

1-*O*-(13-(4-Benzoylphenyl)-9-(*Z*)-tridecenyl)-2-*O*-methoxymethyl-sn-glycerol (34). Compound 34 was prepared in 86% yield using the procedure described to prepare compound 27: 1 H NMR δ 1.24–1.39 (m, 8H), 1.52–1.60 (m, 2H), 1.68–1.77 (m, 2H), 1.97–2.04 (m, 2H), 2.06–2.13 (m, 2H), 2.70 (t, 2H, J = 7.6 Hz), 3.42 (s, 3H), 3.40–3.80 (m, 7H), 4.75 (s, 2H), 5.34–5.45 (m, 2H), 7.27–7.82 (m, 9H); 13 C NMR δ 26.0, 26.7, 27.3, 29.2, 29.4, 29.4, 29.6, 29.7, 31.1, 35.5, 55.6, 63.7, 71.1, 71.8, 78.5, 96.6, 128.2, 128.3, 128.9, 129.9, 130.3, 130.7, 132.1, 135.1, 137.9, 147.8, 196.5; HR-MS [MNa⁺] m/z calcd for $C_{31}H_{44}O_{5}$ Na 519.3081, found 519.3079.

1-*O*-Methoxymethyl-2-*O*-(13-(4-benzoylphenyl)-9-(*Z*)-tridecenyl)-sn-glycerol (35). The same method used to prepare 34 was used to prepare 35 in 88% yield: 1 H NMR δ 1.22–1.40 (m, 8H), 1.52–1.77 (m, 4H), 1.95–2.13 (m, 4H), 2.70 (t, 2H, J = 7.6 Hz), 3.37 (s, 3H), 3.46–3.77 (m, 7H), 4.63 (s, 2H), 5.34–5.45 (m, 2H), 7.27–7.82 (m, 9H); 13 C NMR δ 26.1, 26.7, 27.3, 29.2, 29.4, 29.5, 29.7, 30.0, 31.1, 35.5, 55.3, 62.6, 67.0, 70.3, 78.5, 96.7, 128.2, 128.3, 128.9, 129.9, 130.3, 130.7, 132.2, 135.1, 137.9, 147.8, 196.5; HR-MS [MNa⁺] m/z calcd for C₃₁H₄₄O₅Na 519.3081, found 519.3095.

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Supporting Information Available: Procedures for enzymatic incorporation of [32P]-phosphate into the benzophenone-containing analogues, for covalent modification of plasma proteins, and for the preparation of compounds **9** and **10** and ¹H and ¹³C NMR spectra for all new compounds and intermediates. This material is available free of charge via the Internet at http://pubs.acs.org. JO052030W